



Research Article

ISOLATION AND CHARACTERIZATION OF INCENSOLE AND DA-23-IV (NEPHTHENOL) FROM ETHYL ACETATE STEM BARK EXTRACT OF PSEUDOCEDRELA KOTSCHYI SCHEINF HARMS (MELIACEAE)

^{1,*} Dafam, D.G., ²Agunu, A., ³Ahmed, A.H.M., ²Ibrahim, H., ¹Kagaru, D.C., ¹Ohemu, T.L., ¹Okwori, V. A., ¹Olotu, P.N., ²Ilyas, N., ⁴Agwom, F. M., ⁵Ogira, J.O. and ⁶David, J.

¹Department of Pharmacognosy and Traditional Medicine, University of Jos, Jos-Nigeria

²Department of Pharmacognosy and Drug Development Ahmadu Bello University, Zaria, Nigeria

³Department of Chemistry and Natural Products, University of Western Cape, South Africa

⁴Department of Pharmaceutical and Medicinal Chemistry, University of Jos, Jos, Nigeria

⁵Department of Pharmaceutical and Medicinal Chemistry, Bingham University Karu, Nasarawa, Nigeria

⁶Department of Medical Biotechnology, Biotechnology Development Agency, Abuja, Nigeria

Received 15th September 2023; Accepted 24th October 2023; Published online 30th November 2023

Abstract

Pseudocedrela kotschy Schweinf Harms (Meliaceae) is used in ethnomedicine to treat various disease conditions (like ulcers, sores, rheumatism, leprosy, syphilis, fevers and tumors). The dried pulverized bark material was extracted using maceration method with methanol. The methanolic extract obtained was fractionated into water, hexane and ethyl acetate fractions. Repeated Column chromatography and preparative thin-layer chromatography (TLC) of the ethyl acetate fraction yielded two pure compounds, incensole and nephtenol. The structures of the isolated compounds were elucidated using nuclear magnetic resonance spectroscopy (1D and 2D ¹HNMR and ¹³CNMR). This is the first time report of Incensole a cembrane diterpenoid compound in the stem bark of the plant *P. kotschy*.

Keywords: *Pseudocedrela kotschy*, Incensole, Nephtenol, Chromatography.

INTRODUCTION

Pseudocedrela kotschy Schweinf Harms (Meliaceae) stem bark was observed to have rough texture with brittle and fibrous fracture. The taste is bitter and when dried, pieces assumed curved convex shape. The powder form of the bark is brown to white in colour (Dafam *et al.*, 2019). *P. kotschy* has numerous uses in traditional medicine, particularly its bark, roots and leaves. Bark decoctions or macerations are applied externally to ulcers, sores, rheumatism, leprosy, syphilis, yaws, itch, caries and gingivitis. Internally they are used to treat fever, stomach-ache, diarrhoea and dysentery, and as a diuretic and aphrodisiac (Burkill, 1997). In Nigeria the stem bark is used in mixtures to treat trypanosomiasis in livestock, and leaves are administered in veterinary medicine against intestinal worms (Burkill, 1997). The Tarok speaking communities use the bark and leaves to treat fevers, pains/inflammation and treatment of tumors in the form of powder and decoction (Dafam *et al.*, 2016). The bark yields a brownish dye that has been used in West Africa for dyeing cloth. The plant is occasionally planted as an ornamental shade tree and roadside tree. In Nigeria the leaves are used as a green manure (Burkill, 1997). Phytochemical screening of the plant revealed the absence of anthraquinones and cardiac glycosides, whereas saponins, tannins, flavonoids, steroids and terpenes were shown to be present (Dafam *et al.*, 2018;). The dental cleansing effect (Akande and Hayashi, 1998), antiepileptic (Anuka *et al.*, 1999), anti-inflammatory activity (Otimenyi *et al.*, 2004), analgesic (Musa *et al.*, 2005), anticonvulsant (Odugbemi, 2006), of the plant have been reported. Okunade *et al.*, (2007) also reported its dental cleansing effects.

In addition, the plant has other biological effects such as antibacterial (Koné *et al.*, 2004), antimalarial (Asase *et al.*, 2005; Dawet and Yakubu, 2014), antipyretic (Akuodor *et al.*, 2013), heamatinic, antidiabetic activities and protective effect on the renal system (Ojewale *et al.*, 2014; Georgewill and Georgewill, 2009; Bothon *et al.*, 2013.,) and antiproliferative activities of the root (Olakunle *et al.*, 2015).

MATERIALS AND METHODS

Experimental

General procedures. Thin-layer Chromatography (TLC) was carried out on Si gel 60 F254 Merck®. Accelerated Gradient Chromatography - AGC (Bäckström, 1993), a form of Medium Pressure Liquid Chromatography (MPLC) was carried out on columns packed with Si gel 60, 0.040- 0.063mm Merck®. The MPLC workstation was from University of Western Cape, South Africa. NMR was carried out on a Bruker 400 MHz spectrometer.

Collection and preparation of Plant Material

Stem bark of *Pseudocedrela kotschy* was collected from Langtang North local government area of Plateau state, Nigeria in August, 2014. Photographs of the freshly collected plant were taken in its natural habitat. The plant was identified by comparing with voucher specimens deposited at the Herbaria of the Federal College of Forestry Research Institute Jos, and authenticated in the Department of Botany and plant Sciences, Ahmadu Bello University Zaria, Nigeria by the taxonomist, Mr. Mohammed Musa and a voucher specimen number, ABU 960243 was given. The stem bark of the plant collected was sliced into pieces to facilitate its drying under the shade.

*Corresponding Author: Dafam, D.G.,
Department of Pharmacognosy and Traditional Medicine, University of Jos,
Jos-Nigeria.

Grinding was carried out using a grinding machine to powder form and stored in an airtight container to prevent it from microbial attack. This was preserved according to methods described in the African Herbal Pharmacopoeia (AfrHP) (2010) and African Pharmacopoeia (1985).

Extraction and Isolation

The dried, powdered stem bark of *P. kotschyi* was weighed (800 g) and extracted using maceration method with 3.5 Litres of methanol for 20hrs at room temperature, with occasional shaking and the extract was filtered. The filtrate was concentrated under reduced pressure using rotavapor and the percentage yield determined. The methanol extract (146.30 g) was separately partitioned with water, hexane and ethyl acetate hexane (500 mL) and the aqueous portion (fraction) containing the hexane was concentrated to dryness using rota vapor at a temperature of 65°C. The aqueous portion was further partitioned with ethyl acetate (400 mL) and distilled water (100 mL) in a separating funnel and shaken. The aqueous layer was collected and concentrated using a rotavapor and later on a water bath. The amount of yield obtained was determined and kept for use when needed. The dry-pack method of column chromatography (CC) was adopted. The column (4 x 60) cm was packed with silica gel. The extract was mixed with an equivalent amount of silica and was initially pre-absorbed with Dichloro methane (DCM) was allowed to dry and added to the previously packed adsorbent in the column. The column was gradually eluted with different solvent system; 100 % Hexane, (Hexane/EtOAc) in the ratio (9:1; 8:2; 7:3; 3:2; 1:1; 2:3; 3:7; 2:8) and 100 % ethyl acetate. The TLC was carried out for the various fractions collected and was developed in different solvent systems in varying concentrations. The ethyl acetate fraction was profiled using Hexane/EtOAc (3:2) and the developed chromatograms were observed under UV light at 256 and 366 nm wavelengths and spread with vanillin sulphuric acid for the spots separation visibility when heated in an oven gradually up to 105°C. On repeated CC and TLC analysis of the combined fractions eluted with ethyl acetate, pure fraction coded DA-23-I (Incensole) showed chromatograms of single spot signifying a state of purity of the isolated compound as colourless oil characteristic of incensole (21.13 g /40.50 mg). and fraction coded DA-23-IV (Nephtenol) also showed single spot on TLC in different solvent system.

Characterization and Structure Determination of the Isolated Compound

The structures of the isolated compounds were elucidated through spectroscopic methods to include ultraviolet (UV), infra-red spectrometry (IR), hydrogen nuclear magnetic resonance ¹H(NMR) and ¹³CNMR. The most useful data concerning the chemical structures were furnished by 1D and 2D NMR spectroscopy. From 1H NMR, JMOD, 1H-1H COSY, HSQC and HMBC experiments, the constitution of the compound was determined with the aid of the NOESY spectra, the relative configurations were elucidated. In the isolated compounds, the numbers of asymmetric carbons were determined, and they were all characterized stereochemically. NMR studies allowed complete 1H and 13C assignments for the characterization of the compound.

RESULTS AND DISCUSSION

Incensole from Ethyl acetate fraction of the stem bark of *P. kotschyi*

The compound was isolated as thick oil. The NMR data supported a diterpene skeleton of cembrane type. Signals from ¹H NMR showed two olefinic protons at 5.04, 5.06 ; hydroxylated proton at 3.27 (d, 9.7 Hz); two high field shift methyls at 1.48, 1.60 (both singlet), each attached to a double bond; methyl signal at 1.04 (s); signal for isopropyl group [two doublet methyl signals at 0.887, 0.889 (J = 7.9 Hz); and a proton at 1.95 (m)] these were supported by 1H-1H Cosy spectra which showed a cross peak between the two methyls and the proton. ¹³C NMR spectra showed 20 signals of 20 carbons which support the diterpene skeleton. Those signals could be splitted using DEPT-135 into 5 methyls (0.887, 889, 1.04, 1.48, 1.60), 7 methylene (24.8, 30.6, 30.7, 32.4, 33.7, 36.4, 38.6), 4 methines, two of them are olefinic and one hydroxylated (34.8, 75.6, 121.8, 125.1) and 4 quaternary carbons (84.2, 88.6, 134.2, 134.3). Furthermore, the HMBC spectra showed correlations between (δ_H/δ_C): Me's 0.887, 0.889, 5.06/88.6; Me 1.04/84.2; 1.48/121.8, 134.3 and 1.60/125.1, 134.2. The long range C-H correlation deduced from the COLOC spectrum, H-H COSY coupling and NOESY, prove that compound is the macrocyclic diterpenoid: 1,12-epoxy-3,7- cembradien-11-ol, known as incensole.

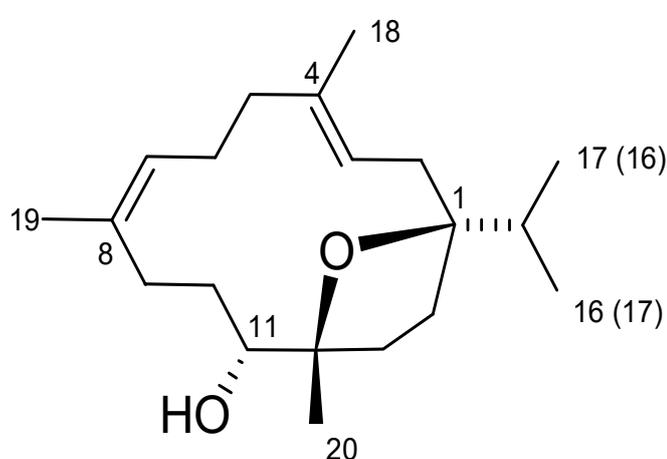
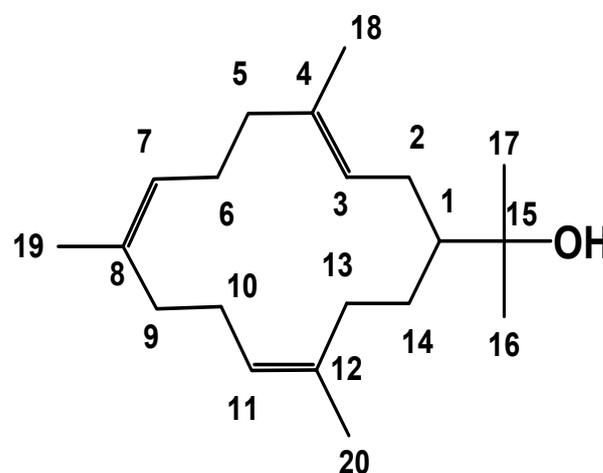
Table 1. Summary of NMR Data on DA-23-I, of Ethyl acetate fraction from the stem bark of *P. kotschyi*

| Position | δ_C (ppm) | DEPT | δ_H (ppm)/HSQC | H-H COSY | HMBC |
|----------|------------------|-----------------|-----------------------|----------|-----------------------|
| 1 | 88.53 | C | | | H-2;H-14;H-16/17;H-13 |
| 2 | 33.62 | CH ₂ | 2.00 | H-3 | H-14;H-15 |
| 3 | 121.74 | CH | 5.08 | H-2 | H-18;H-5;H-14;H-15 |
| 4 | 134.16 | C | | | H-2;H-6;H-18H-5 |
| 5 | 34.79 | CH ₂ | 1.88 | H-6 | H-18;H-3;H-6 |
| 6 | 24.8 | CH ₂ | 1.48 | H-5;H-7 | H-7;H-5;H-9 |
| 7 | 125.10 | CH | 5.05 | H-6 | H-5;H-19;H-9 |
| 8 | 134.23 | C | | | H-10;H-5;H-6;H-11 |
| 9 | 32.31 | CH ₂ | 1.85 | H-10 | H-7;H-19;H-11 |
| 10 | 36.32 | CH ₂ | 1.89 | H-9;H-11 | OH;H-11; |
| 11 | 84.12 | CH | | H-10 | H-20;H-9;H-10;OH |
| 12 | 77.31 | C | | | H-20;OH;H11;H-14;H-10 |
| 13 | 30.68 | CH ₂ | 1.34 | H-14 | H-20;H-11 |
| 14 | 30.59 | CH ₂ | 1.33 | H-13 | H-15;H-2;H-13 |
| 15 | 38.58 | CH | 2.04 | H-16/17 | H-14;H-2;H-3;H-16/17 |
| *16 | 17.96 | CH ₃ | 1.27 | H-15 | H-2;H-14 |
| *17 | 18.04 | CH ₃ | 1.27 | H-15 | H-2;H-14 |
| 18 | 20.63 | CH ₃ | 1.29 | H-3 | H-3;H-5;H-6 |
| 19 | 18.14 | CH ₃ | 1.30 | H-7 | H-7;H-9; |
| 20 | 16.10 | CH ₃ | 1.32 | H-11 | H-11;H-13; |

N.B.: * assignments are interchangeable.

Table 2. NMR data for Nephthenol, of ethyl acetate fraction from the stem bark of *p. kotschy*

| Position | δ_c (ppm) | DEPT | δ_H (ppm)/ HSQC | H-H COSY | HMBC |
|----------|------------------|-----------------|------------------------|----------|----------------------|
| 1 | 48.47 | CH | 1.54 | H-2;H14 | |
| 2 | 28.45 | CH ₂ | 1.58 | H-1;H-3 | H-14;H-16/17;H-4 |
| 3 | 125.96 | CH | 5.07 | H-2 | H-1;H-18 |
| 4 | 133.35 | C | | | H-18;H-5;H-3;H 6;H 2 |
| 5 | 38.81 | CH ₂ | 2.12 | H-6 | H-3;H-7 |
| 6 | 24.67 | CH ₂ | 2.08 | H-5;H-7 | H-3;H-18;H-19 |
| 7 | 125.78 | CH | 4.98 | H-6 | H-9;H-19;H-5;H-6 |
| 8 | 133.03 | C | | | H-10;H-19;H-9 |
| 9 | 39.39 | CH ₂ | 2.09 | H-10 | H-20;H-7 |
| 10 | 24.02 | CH ₂ | 2.11 | H-9;H-11 | H-7;H-13;H-20 |
| 11 | 125.00 | CH | 4.97 | H-10 | H-9;H-14;H-20;H-13 |
| 12 | 134.02 | C | | | H-14;H-1;H-10;H-20 |
| 13 | 37.42 | CH ₂ | 1.65 | H-14 | H-11;H-1 |
| 14 | 28.26 | CH ₂ | 1.54 | H-13;H-1 | H-2 |
| 15 | 73.96 | C | | | H-16/17;H-14;H-2;OH |
| 16 | 27.67 | CH ₃ | 1.29 | | OH;H-17;H-1 |
| 17 | 27.51 | CH ₃ | 1.29 | | OH;H-16;H-1 |
| 18 | 29.71 | CH ₃ | 1.61 | | H-3;H-5 |
| 19 | 15.32 | CH ₃ | 1.61 | | H-7;H-9 |
| 20 | 15.57 | CH ₃ | 1.61 | | H-11;H-13 |

Fig. 1. The structure of Incensole from the ethyl acetate fraction of the stem bark of *P. kotschy*Fig. 2. The depicted structure of Nephthenol from ethyl acetate fraction of the stem bark of *P. kotschy*

The isolated compound is soluble in dichloro methane (CH₂Cl₂) and Ethyl acetate (EtOAc). The ¹H NMR data for this compound revealed that it is in agreement with that obtained by previous researchers (Corsano and Nicoletti, 1967), who isolated and characterized the same compound, incensole, from the neutral fraction of frankincense, produced by *Boswellia carteri*. Its chemical synthesis (Kato, 1976) and ¹³C NMR spectral data (Gacs-Baitz, *et al.*, 1978) have also been reported. It is to be noted that this is the first time Incensole is reported in *P. kotschy* Schweinf Harms (Meliaceae). The second isolated compound showed similar NMR data with incensole, which indicated a cembrane diterpene derivative. ¹H NMR showed three olefinic protons signals at 5.07, 4.98, and 4.97 (br t, J=6.1 Hz), 5.11 (br t, 6.0 Hz); 5 methyl singlet signals 1.19 (X₂), 1.55, 1.56 (X₂). ¹³C NMR spectra showed 20 signals of 20 carbons which support the diterpene skeleton. This was splitted with DEPT-135 which showed five methyls, (27.67, 27.51, 29.71, 15.32 and 15.57), seven methylenes (28.45, 38.84, 24.67, 39.43, 24.02, 37.42, 28.26), four methines (48.47, 125.96, 125.78, and 125.00).and four quaternary carbons (133.35, 133.03, 134.02 and 73.93). The HMBC showed correlations between (δ_H/δ_C): 1.58/28.45, 2.12/38.84, 2.08/24.02, 4.98/125.78, 2.09/39.43 1.65/37.42, 1.54/28.26 and 2.11/23.99. The data indicated cembrane diterpene with three double bonds and oxygenated isopropyl group.

REFERENCES

- African Pharmacopoeia (1985). Pharmacopoe Africaine OAU/STR Scientific Publication. Prepared by Inter African Committee on Medicinal Plants and Akande, J.A. and Hayashi, Y.1998. Potency of extract contents from selected tropical chewing sticks against *Staphylococcus aureus* and *Staphylococcus auricularis*. *World Journal of Microbiology and Biotechnology*, 14:235-238.
- Anuka, J.A., Ijezie, D.O., and Ezebnik, O.N. (1999). Investigation of pharmacological actions of the extract *Pseudocedrela kotschy* in laboratory animals. Abstracts of the Proceedings of XXV 11th Annual Regional Conference of WASP, pp: 9-10.
- African Traditional Medicine. 1st Ed. Vol.1 Lagos-Nigeria.
- Asase, A., A.A., Oteng-Yeboah, Odamtten, G.T., and Simmonds, M.S.J. (2005). Ethnobotanical study of some Ghanaian anti-malarial plants. *Journal of Ethnopharmacology*, 99: 273-279.
- Bothon, F.D.A., E. Debiton, F. Avlessi, C. Forestier, C. Teulade and K.C. Sohounhlone, 2013. In vitro biological effects of two anti-diabetic medicinal plants used in Benin as folk medicine. *BMC Complementary Alternative Medicine*. 13(51): 1- 8.
- Burkill, H.M. (1997). The useful plants of West Tropical Africa. 2nd Edition. Volume 4, Families M-R. Royal

- Botanic Gardens, Kew, Richmond, United Kingdom. Pp 969
- Dafam, D.G., Agunu, A., Ibrahim, H., Ojerinde, O.S., Ohemu, T.L., Okwori, V.A., Olotu, P.N. and Ilyas, N (2018). Phytochemical screening and antioxidant activity of *Pseudocedrela kotschy* Schweinf Harms (Meliaceae) and *Strophanthus sarmentosus* DC (Apocynaceae). *Journal of Pharmacy and Bioresources* 15(1) 78-83
- Dafam, D.G., Agunu, A., Ibrahim, H., Ajima, U., Ohemu, T.L., Okwori, V. A., Kagaru, D.C. and N. Ilyas (2016). A Preliminary ethno-medical survey of plants used in the traditional management of cancer and related diseases amongst Tarok people of Plateau state, north-central Nigeria. *European Journal of Medicinal Plants*, 16 (4): 1-10.
- Dawet, A and. Yakubu, D.P. (2014). Antiplasmodial Efficacy of Stem Bark Extracts of *Pseudocedrela kotschy* in Mice Infected with *Plasmodium berghei*. *British Journal of Pharmaceutical Research* 4(5): 594-607, 2014
- Gacs-Baitz E., Radics L., Fardella G. and Corsano S. (1978): Carbon-13 nuclear magnetic resonance studies of some fourteen-membered macrocyclic diterpenes; *J. Chem. Res. (S)*, (4), 146.
- Georgewill, O.U. and Georgewill, A.O. (2009). Effects of extract of *Pseudocedrela kotschy* on blood glucose concentration of Alloxan-induced diabetic rats. *Eastern Journal of Medicine*. 14(1): 17-19.
- Kato T. (1976). Synthesis of D,L-incensole; *Chem. Lett.*, 1191.
- Koné, W.M., Atindehou, K.K., Terreaux, C., Hostettmann, K., Traore, D and Dosso, M. (2004). Traditional medicine in North Côte d'Ivoire: Screening of 50 medicinal plants for antibacterial activity. *Journal of Ethnopharmacology*, 93: 43-49.
- Musa, Y.M., Haruna, A.K., Ilyas, M., Yaro, A.H., Ahmadu, A.A., and Usman, H. (2005). Analgesic and anti-inflammatory activities of the leaves of *Pseudocedrela kotschy* Harms (Meliaceae). Books of abstracts of the 23rd National Scientific Conference of the Nigerian Society of Pharmacognosy, Pp: 88-89.
- Ojewale, A.O., Adekoya, A.O., Faduyile, F.A., Yemitan, O.K., and Odukanmi, A.O. (2014). Nephroprotective Activities of Ethanolic Roots Extract of *Pseudocedrela kotschy* against Oxidative Stress and Nephrotoxicity in Alloxan-induced Diabetic Albino Rats. *British Journal of Pharmacology and Toxicology* 5(1): 26- 34,
- Okunade, M.B., Adejumbi, J.A., Ogundiya, M.O. and Kolapo, A.L. (2007). Chemical, phytochemical compositions and antimicrobial activities of some local chewing sticks used in South Western Nigeria. *Journal of Phytopharmaceutical and Natural Products*. 1(1): 49-52.
- Olakunle, O. K., Robert, L. C., Hilaire, M. K., Sergei, N., Kwashie, A.A. and Yasmine, M.K, (2015). Antiproliferative Activities of *Fagaraxanthoxyloides* and *Pseudocedrela kotschy* Against Prostate Cancer Cell Lines. *International Journal of Cancer Research and Treatment*. 35 (3):1453-1458
- Otimenyi, S.O., Uguru, M.O., and Atang, (2004). Anti-inflammatory and Analgesic activities of *Ficus thoningii* and *Pseudocedrela kotschy* extracts. *Nigerian Journal of Pharmaceutica Research*. 3 (1):82-85.

SUPPLEMENTARY DATA

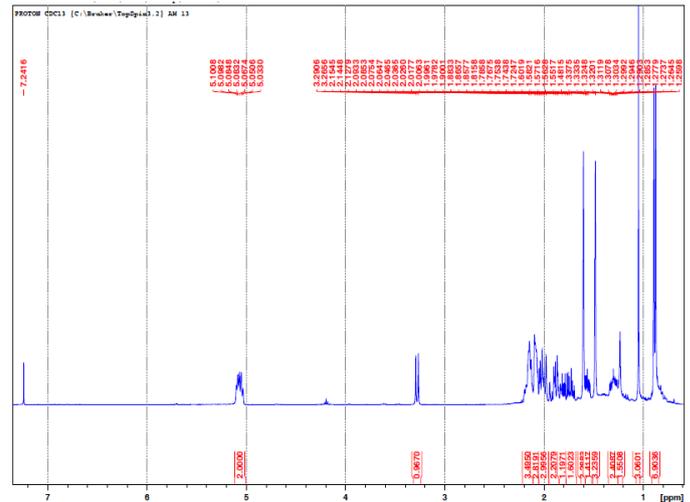


Fig. 1: $^1\text{H-NMR}$ spectra of Icencole from the ethyl acetate fraction of the stem bark of *P. kotschy*.

DA_23_1_C1

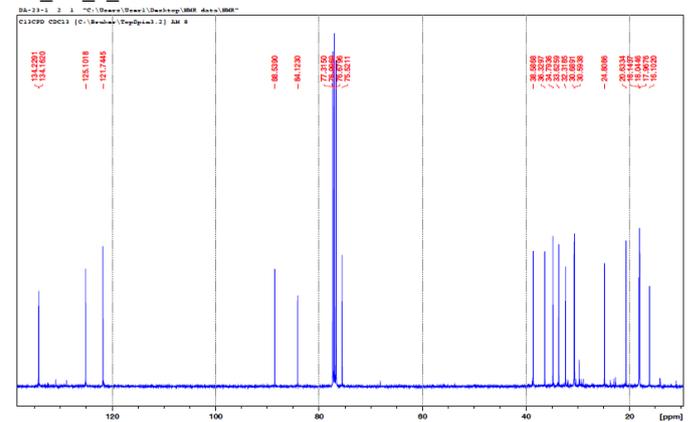


Fig. 2. $^{13}\text{C-NMR}$ spectra of Icencole from the ethyl acetate fraction of the stem bark of *P. kotschy*.

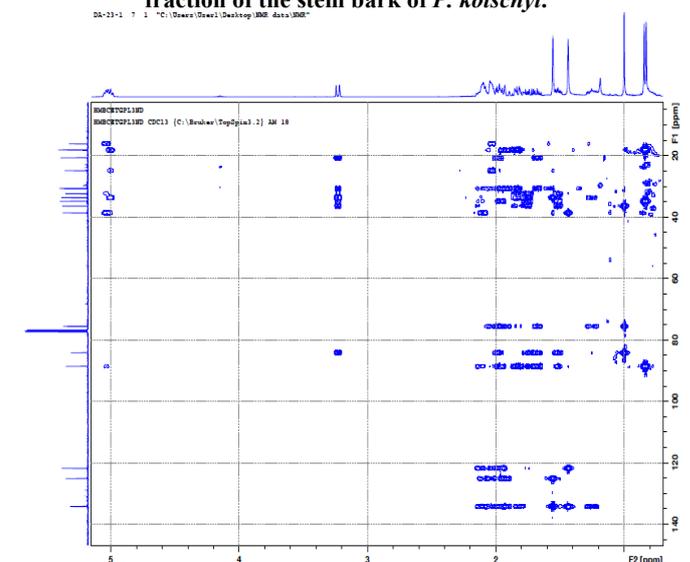


Fig. 3. The HMBC1 spectra of Icencole from the ethyl acetate fraction of the stem bark of *P. kotschy*.

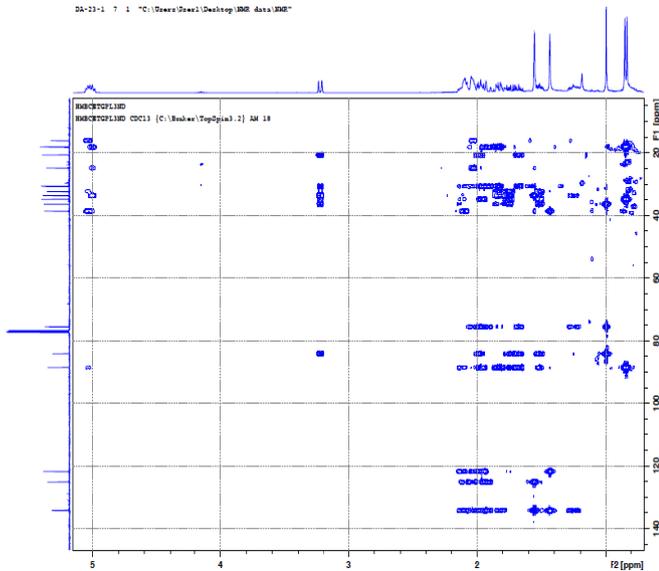


Fig. 4. The HSQC1 spectra of Icencole from the ethyl acetate fraction of the stem bark of *P. kotschy*.

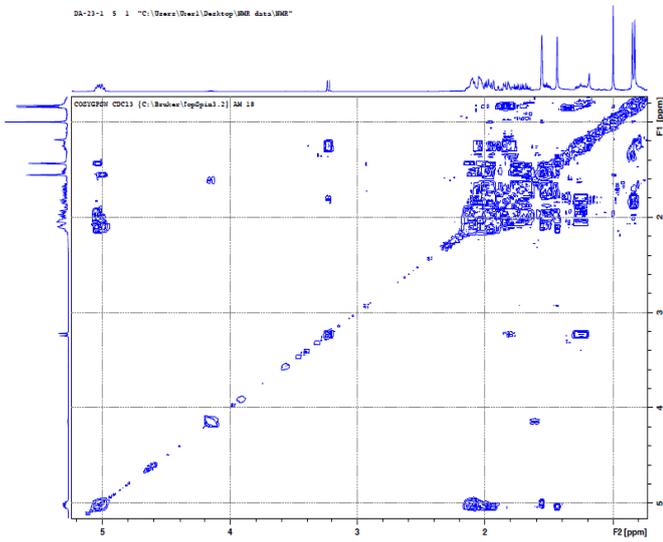


Fig 5. The COSY1 spectra of Icencole from the ethyl acetate fraction of the stem bark of *P. kotschy*.

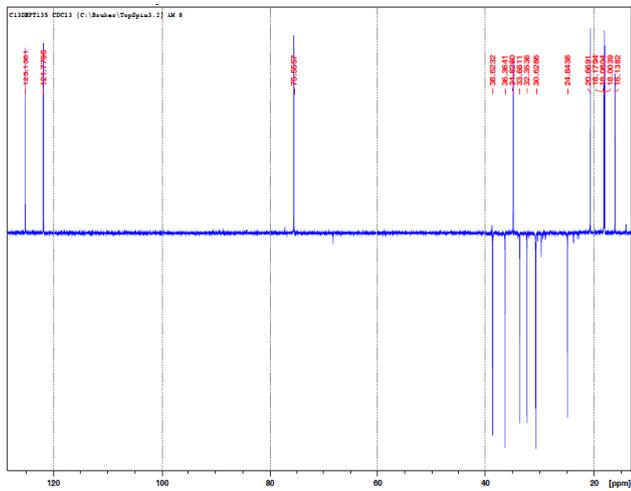


Fig 6. The DEPT1 of Icencole from the ethyl acetate fraction of the stem bark of *P. kotschy*.

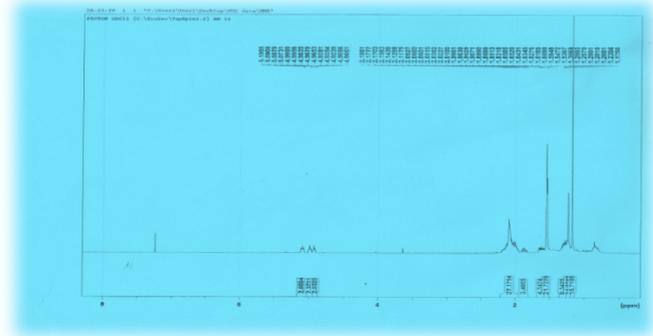


Fig. 7. The ¹H NMR spectra of Nephthenol from the ethyl acetate fraction of the stem bark of *P. kotschy*.

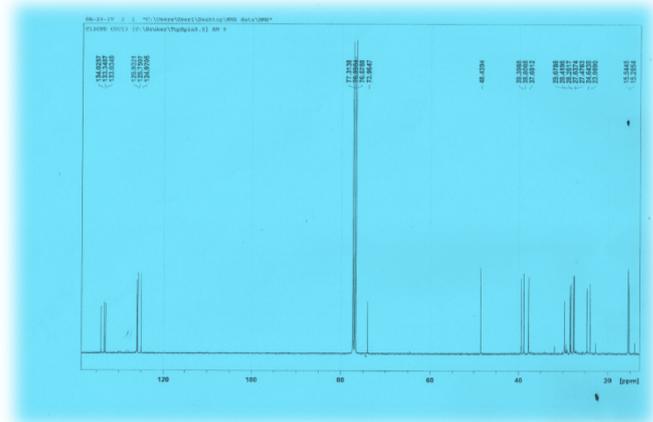


Fig. 8. ¹³C NMR spectra of Nephthenol from the ethyl acetate fraction of the stem bark of *P. kotschy*.

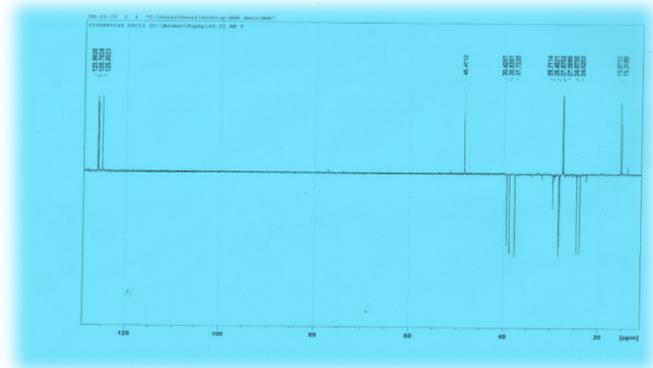


Fig. 9. The DEPT spectra of nephthenol from the ethyl acetate fraction of the stem bark of *P. kotschy*.

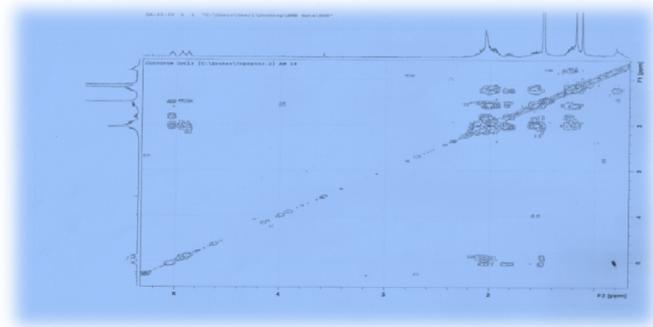


Fig. 10. The COSYGPSW spectra of Nephthenol from the ethyl acetate fraction of the stem bark of *P. kotschy*.

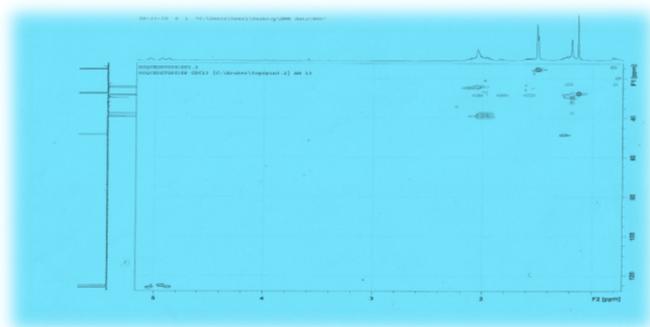


Fig.11. The HSQC spectra of Nephthenol from the ethyl acetate fraction of the stem bark of *P. kotschy*.

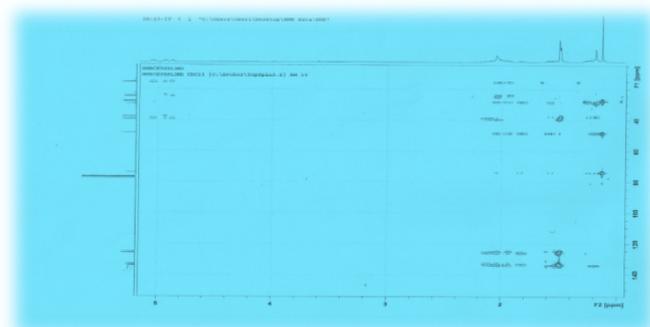


Fig.12. The HMBC spectra of nephthenol from the ethyl acetate fraction of the stem bark of *P. kotschy*.
