

MICROBIAL MODULATION OF NEUROINFLAMMATION AND NEURODEGENERATION

*Shivani Kale, Snehal Kadam, Geeta Shinde, Aman Upaganlawar, Manoj Maajan and Chandrashekhar Upasani

Department of Pharmacology, SNJBs Shriman Sureshdada Jain College of Pharmacy, Chandwad, Nashik, India

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Abstract

Microbial communities significantly produce neuroinflammation and neurodegeneration, which are attributes of neurological diseases such as multiple sclerosis, parkinson’s disease, and alzheimer’s disease. The gut-brain axis (GBA) serves as a significant barrier, with gut microbiota modulating microglial activation, cytokine production, and blood-brain barrier integrity. Specific microbial taxa, including Firmicutes and Bacteroidetes, produce pro-inflammatory metabolites, regulating the immune system and exacerbating neuroinflammation, while pathogenic microbes like Herpesviruses and Chlamydia contribute to neurodegeneration. Understanding this complex interplay has profound implications for therapeutic intervention and disease management. This offers promising avenues via microbiome-targeted therapies, novel biomarkers, and elucidation of the microbiome-neuroinflammation axis, underscoring the need for further investigation into microbial modulation to develop innovative strategies against neurological disorders.

Keywords:Microbiome, Neuroinflammation, Neurodegeneration, Gut-brain axis, Microglial activation.

INTRODUCTION

In a complicated and varied ecosystem, microbiota is mainly made up of bacteria, but it also includes archaea, fungi, viruses, and protozoa (1). These microorganisms are crucial for maintaining human health as they regulate host immunity while maintaining the intestinal mucosal barrier (2). Multiple processes including the immune system and the vagus nerve interact with bioactive molecules, and the endocrine system, that facilitate the microbiota neurological interface (3). The brain and microbiota interact with each other through the release of neuroactive compounds, the activation of the gut immune system, and the network of cells in the enteric nervous system (ENS), connecting the gut to the central nervous system (CNS) via the vagus nerve. In particular, the gut microbiota creates bioactive molecules like neuropeptides, neurotransmitters hormones, and neuropeptides that influence the enteric and systemic nervous systems with an effect on behaviour and brain function(4).

Types of intestinal micro biota

The sterile fecal gut undergoes rapid colonization with micro biota during the perinatal period, primarily through exposure to maternal vaginal and fecal flora, as well as environmental microorganisms. As we grow older, the variety and amount of good bacteria in our gut increase. Scientists group these bacteria by their characteristics, shape, and function. Research shows that most gut bacteria come from four main groups *Formicates*, *Bacteroidetes*, *Proteobacteria*, and *Actinobacteria*. Each person's gut bacteria are unique, like fingerprints. The mixture of bacteria differs from person to person, and their roles in keeping us healthy also vary (5).

The main gut bacteria groups are listed below with a suitable example are listed below: (Figure 1).

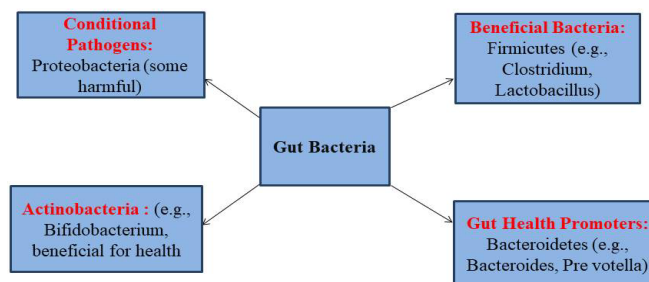


Figure 1. These groups are presented as connected to a central gut bacteria node, indicating their role within the gut microbiome

Gut-brain axis: More than 60 years ago, scientists noticed a relationship between intestinal microbes and their metabolic products and hepatic coma. Furthermore, they found that antibiotics could decrease symptoms of hepatic encephalopathy (6) (Figure 2).

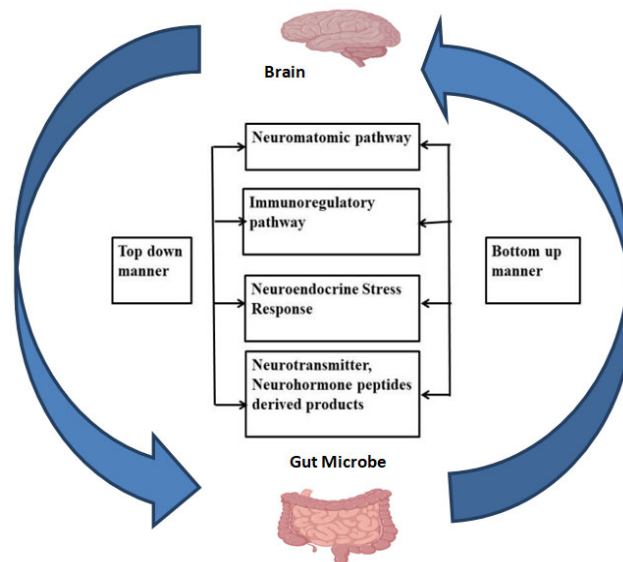


Figure 2. Mechanism of action of inflammation Correlation of Gut Brain axis

*Corresponding Author: Shivani Kale, Department of Pharmacology, SNJBs Shriman Sureshdada Jain College of Pharmacy, Chandwad, Nashik, India.

The human brain and gut are connected to each other. The GBA acts as a bidirectional communication between the CNS and the body's ENS. The ENS, which spans from the bottom third of the esophagus to the rectum, is an intricate structure of sensory, motor, and interneuron cells embedded in the gastrointestinal system's wall (7,8). It links the brain's emotional and cognitive centers to the normal operation of the peripheral bowel through indirect and direct routes (9,10).

Microbial Influence on Host Health

1. The gut microbiome influences neuroinflammation through cytokine production, immune cell modulation, and blood-brain barrier permeability (Figure 3).

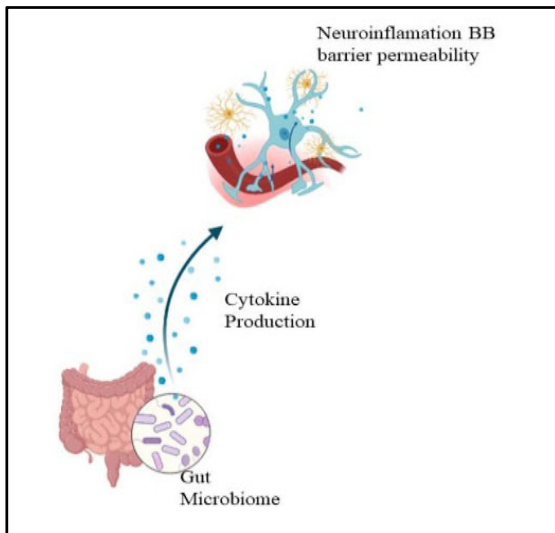


Figure 3. Gut microbiota Influence Neuroinflammation

2. Microbiota-generated metabolites modulate neuroinflammatory responses and neurodegenerative processes (11) (Figure 4).

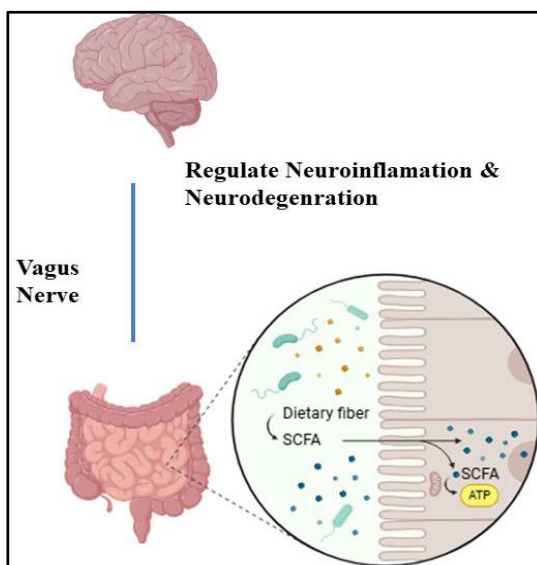


Figure 4. Microbiome Regulates the Neuroinflammation and Neurodegeneration

3. Microbiome changes, known as dysbiosis, play a role in the development of neurological degenerative conditions, including Alzheimer's disease (AD), Parkinson's disease (PD), and Multiple sclerosis (MS)(12).

4. Microbiome modulation regulates neuroinflammation through vagal afferent signaling, gut-brain axis homeostasis, and reciprocal microbiome-gut-brain interactions (13).

Neuroinflammation

Neuroinflammation is a silent contributor to various metabolic disorders. This process involves immune cells protecting the brain producing proinflammatory cytokines, chemokines, and small-molecule messengers (14) (Figure 5).

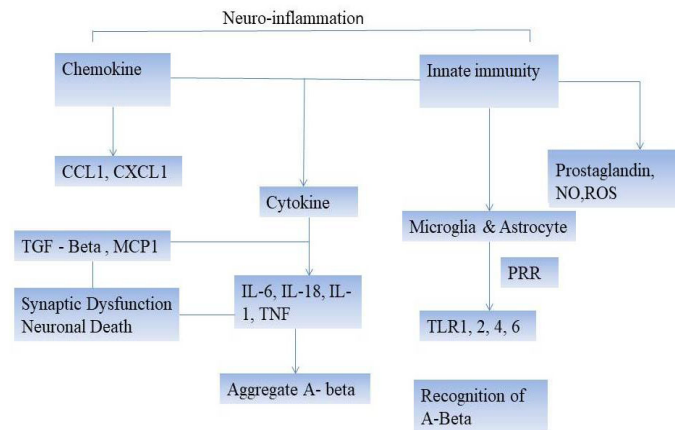


Figure 5. Microglia and astrocytes release inflammatory mediators in that cytokines (IL-1, IL-6, IL-18, TNF), chemokines (CCL1, CCL5, CXCL1), and micromolecules (prostaglandins, NO, reactive oxygen species)

1. Glial activation and cytokine production triggered by amyloid- β ($A\beta$) aggregates.
2. Microglia internalizes $A\beta$ plaques, linking neuroinflammation to AD.
3. Disruption of the cerebral vascular barrier permits vascular endothelium and invading blood cells to contribute to neuroinflammation (15).
4. Pattern recognition receptors on microglia (e.g., TLR1, TLR2, TLR4, TLR6, CD14, CD47, and CD36) recognize $A\beta$ species, triggering inflammatory signaling cascades (16).

In AD, neuroinflammation produces reduced neurogenesis, synaptic dysfunction, and neuronal death. Through a spectrum of mechanisms, pro-inflammatory cytokines like IL-1 and TNF cause damage to nerve cells. Pathological assessment, neuroimaging, and biochemical analysis in blood and cerebrospinal fluid (CSF) have all been used in clinical studies to investigate neuroinflammation in AD. Increased levels of cytokines (TNF- α , IL-6, TGF- β , IL-1, IL-18, and IL-12) have been detected in AD patients' blood and CSF. Microglia are activated in early AD stages, promoting $A\beta$ clearance through CD14 and TLR(17).

Neurodegeneration

Neurodegeneration results in the progressive loss of structure or function of neurons, including their death, which can lead to cognitive, motor, and sensory impairments. Elevated levels of cytokines such as IL-6, TNF- α , IL-1, TGF- β , IL-12, and IL-18 are often linked to neurodegeneration because these molecules play a role in neuroinflammation, a key driver of neuronal damage and death in conditions like AD, PD, MS, etc.

Alzheimer's Diseases

a) Amyloid beta accumulation

The sticky protein A β aggregates in the brain to create plaques that harm neurons. Gut connection A β accumulation may be influenced by the gut microbiota, or the bacterial community in your stomach (18). Dysbiosis, or an unhealthy stomach, can lead to inflammation, which can then move to the brain and promote the formation of A β plaques. A β aggregates faster due to the production of toxic chemicals by some gut bacteria, such as TMAO. Inversely, beneficial compounds (such as butyrate) that lower inflammation and safeguard the brain are produced by beneficial gut flora (19).

b) Tau Protein Polymerization

Tau is a protein that often helps in neuronal activity. Tau is harmed, tangled, and disrupted in AD (20, 21). Gut connection Tau tangles are also influenced by the gut bacteria. Inflammation caused on due to an unbalanced gut increases the risk of tau protein damage and tangle formation. Toxins released by harmful gut bacteria can travel to the brain and promote tau damage. On the other hand, butyrate and other compounds produced by healthy gut bacteria may shield tau proteins and lessen the creation of tangles (22, 23).

Pathway leading to decreased cerebral blood flow

A β oligomers cause capillaries to narrow in AD by stimulating endothelin A (ETA) receptors and causing reactive oxygen species (ROS)(24). Elevated ROS and endothelin-1 levels in AD brains support this mechanism. Perivascular macrophages, microglia, and pericytes may generate ROS. ETA receptors, expressed on all pericyte classes, are activated in AD(25). Inflammatory mediators, such as interleukin-1 β , released by activated microglia and astrocytes, may also decrease cerebral blood flow. Reduced CBF arises from mutations in the microglial TREM2, a receptor, a gene associated with AD susceptibility, This can result in the generation of inflammatory mediators (26) (Figure 6)

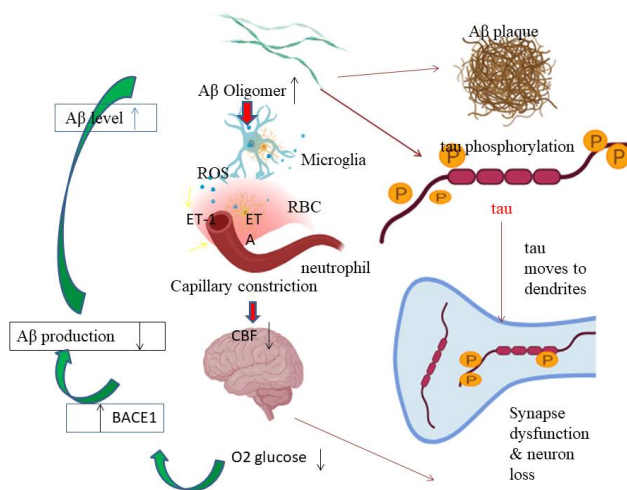


Figure 6. Amyloid Beta and Tau Cascades Diagram

CBF Decrease → Low O₂/Glucose → BACE1 Up-regulation → A β Production

A β Increase → ROS Production → ET-1 Release → Capillary Constriction → CBF Decrease

Parkinson's Diseases

a) Alpha-synuclein misfolding

Dysbiosis can trigger a chronic inflammatory response in the gut. This inflammation can lead to the activation of enteric glial cells and the release of pro-inflammatory cytokines such as IL-6, TNF- α , and IL-1 β . The inflammatory milieu can promote the misfolding of α -synuclein in the ENS. Misfolded α -synuclein can then propagate in a prion-like manner, spreading from the gut to the CNS via the vagus nerve (27). A protein that is often found in the brain, α -synuclein plays a role in the trafficking of synapses. α -synuclein misfolds in PD, forming toxic oligomers and fibrils that eventually combine to form Lewy bodies, a characteristic of the disease. There are multiple ways in which the gut microbiota can affect α -synuclein misfolding. For example, the aggregation of α -synuclein can be modulated by bacterial metabolites including lipopolysaccharides (LPS) and short-chain fatty acids (SCFAs). Gram-negative bacteria's outer membrane contains LPS, which can cause neuroinflammation and encourage α -synuclein misfolding. Conversely, depending on their concentration and type, SCFAs have been demonstrated to have beneficial as well as adverse effects (28).

b) Dopaminergic neuron loss

Dysbiosis of the gut microbiota can trigger inflammation in the gut, leading to the activation of both peripheral and central immune responses. This persistent inflammation in the intestinal lining can contribute to neurological inflammation through the GBA(29). Inflammatory substances circulating in the bloodstream can cross the blood-brain barrier (BBB), allowing inflammatory cytokines to reach the substantia nigra. This process can trigger neuroinflammation and contribute to the degeneration of dopaminergic neurons. Additionally, alterations in gut bacteria with anti-inflammatory properties have been observed in individuals diagnosed with PD (30). A study conducted on individuals with PD revealed that the gastrointestinal mucosa had more *Ralstonia* and less fecal bacteria, and the stool samples had significantly fewer *Blautia*, *Coprococcus*, and *Roseburia species*. Its colon's microbiota had also changed to a more aggressive phase (31). It has been determined bacteria and microorganisms, which are linked with elevated levels of TNF- α and IFN- γ in blood, are relatively abundant in the condition known as Parkinson's disease (PD)(32). This suggests that PD patients have altered gut flora in a systemic sub-inflammatory illness usually, young Pink1 knockout mice can suffer from severe muscle weakness and striatal dopaminergic axon loss when exposed to gram-negative microorganisms, which can lead to minor gastrointestinal problems in the elderly. This indicates that intestinal inflammation and gastrointestinal microbes might interact with a genetic predisposition to PD. Unusually, young Pink1 knockout mice can suffer from severe muscle weakness and striatal dopaminergic axon loss when exposed to gram-negative microorganisms, which can lead to minor gastrointestinal problems in the elderly. This indicates that intestinal inflammation and gastrointestinal microbes might interact with a genetic predisposition to PD (33).

Multiple Sclerosis

Through mechanisms such as increased intestinal permeability, autoimmune responses, compromised long-term inflammation,

and BBB stability an imbalance in gut microbiota contributes to the development and advancement of MS, emphasizing the potential benefits of gut microbiota-targeted therapies (34). MS is a long-term autoimmune condition that impacts the brain and spinal cord. Immune cells damage the protective myelin sheath around neurons, leading to various neurological symptoms. This process involves demyelination, as well as the loss of axons and neurons (35) (Figure 7).

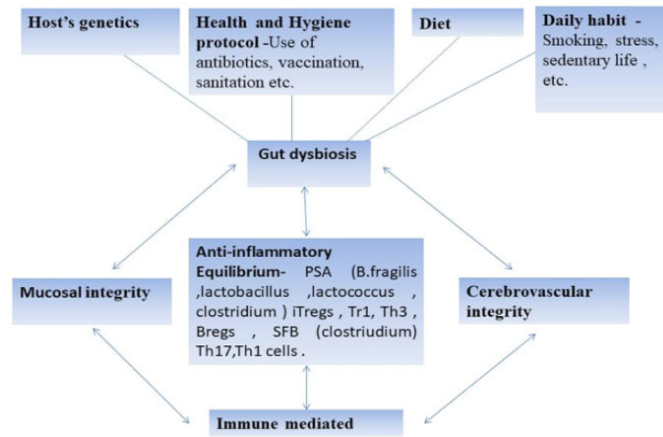


Figure 7. Factors affecting gut dysbiosis

Research suggests a reciprocal link between gut dysbiosis (imbalance) and Inflammation of the brain and spinal cord, particularly in MS. Genetic and external influences, like sanitation, diet, antibiotics, and lifestyle, influence MS risk and also alter gut microbiota composition. Pro-inflammatory dietary factors promote gut dysbiosis, potentially leading to autoimmune CNS diseases. Conversely, anti-inflammatory dietary components support a balanced gut microbiota. Metabolic products, like the Anti-inflammatory properties of SCFAs support gut-brain axis health. The gut microbiome-disease interaction is two-way, requiring further investigation through experimental models and human studies (37). MS is affected by both hereditary and external influences. While its exact nature (neurological or autoimmune) is debated, inflammation and immune dysregulation play key roles. Environmental risk factors include dehydration and electrolyte imbalance. These factors interact with genetic susceptibility, increasing MS risk (38).

Chronic inflammation

MS, chronic inflammation arises from the interaction and coordination of innate and adaptive immune cells, along with the cytokines they release. Activated, CNS-specific cells infected through immunological cells causing oligodendrocyte death, demyelination, and axonal damage (39,40). Th17 cells are key effector cells in MS inflammation, secreting IL-21, IL-17, and IL-22. The gut microbiota modulates immune balance by regulating Treg cell development and Th17 cell maturation, influencing, MS, progression (41).

Autoimmune responses

The development and exacerbation of MS are largely attributed to autoimmune reactions. MS, auto-reactive IgGs and T-cells target myelin components (MBP, PLP, MOG), axonal proteins, and ion channels. Auto-reactive T-cells, potentially generated in gut-associated lymphoid tissue (GALT), play a crucial role (42). The gut microbiota may trigger these T-cells

through molecular mimicry, where microbial components resemble self-antigens. With gut bacteria, the overwhelming majority of vast genetic diversity increases the likelihood of similarity between microbial products (43, 44).

Increases Intestinal permeability

Research suggested that, intestinal permeability increases in approximately 72% of MS patients, which is significantly higher than the 28% rate in healthy controls. By investigations using an immunological model of CNS inflammation, increased intestinal permeability occurs before neurological symptoms appear and further deteriorates as the condition progresses (45). Reduced beneficial agents like SCFAs and anti-inflammatory factors are made up by the gut microbiota, A long side a rise in toxic metabolites and pro-inflammatory cytokines, this can contribute to the breakdown of the intestinal epithelial barrier. As a result, immune cells activated by gut microbiota in the peripheral areas, particularly in GALT, may be further influenced by heightened intestinal permeability (46).

Disruption of the Blood-brain Barrier

For the purpose of maintaining CNS homeostasis, the cerebral barrier regulates the flow of both Substances and cells moving between the bloodstream and the brain (47). However, in MS, BBB breakdown compromises this selective permeability. Disruption occurs early in MS development, enabling leukocyte migration into the CNS and exacerbating disease progression (48,49).

Microbial modulation

a) Lactobacillus Species

Gut-derived GABA, produced by *lactic acid bacteria (LAB)*, regulates the vagus nerve pathway, influencing CNS function (50). Low GABA levels are linked to depression, but LAB strains like *L. reuteri* and *Lactobacillus murine* can restore balance and mitigate symptoms. Additionally, LAB metabolites: Convert L-dopa to dopamine (*E. faecalis/ E. faecium*) (51). Modulate dopaminergic pathways (*L. plantarum* PS128), Upregulate 5-HT receptor 4 expression (52). (*L. rhamnosus* GG, *Ligilactobacillus salivarius Li01*), Decrease serum 5-HT levels (*Ligilactobacillus salivarius Li01*) These findings highlight LAB's role in producing neurotransmitter precursors and modulating host behavior

b) Bifidobacterium Species

Gut microbes contribute to MS disease development and progression. Alterations in gut bacteria composition contribute to immune system dysfunction, inflammation, and neuronal damage (53). Targeting the gut microbiome through probiotics, prebiotics, fecal transplants, and dietary changes offers therapeutic potential (54). Research is exploring how gut bacteria interact with the host to exacerbate or alleviate MS symptoms. Personalized micro biome-based treatments and innovative technologies are being developed

Bifido bacterium exerts beneficial effects on spinal cord injury cell damage:

- Enhance Target Cell Activity
- Safeguarding Gut Health
- Modulating dendritic cells and macrophages
- Suppressing Th2 and Th17 response(55).

Although the exact mechanisms remain complex, recent research highlights Bifido bacterium's surface polysaccharides, proteins, and metabolic products as key mediators of immune homeostasis, promoting tolerance and balance(56).

Probiotic

Therapeutic interventions targeting gut microbiota (GM) and its metabolites offer promise for treating microbiota-related diseases(57). Two potential approaches include manipulating disease progression pathways and utilizing probiotics. Research supports the effectiveness of probiotics, with meta-analyses showing that probiotics consumption improves cognition in AD patients by reducing inflammation and oxidative biomarkers, and also enhances cognitive function when used in intestinal microbiota balance therapy(58)(Table 1).

Table 1. Probiotic and their Effect

Probiotic	Effect	Reference
<i>B breve</i>	Prevented cognitive decline	(59)
<i>Lactobacillus acidophilus</i>	Enhance memory	(60)
<i>Lactobacillus fermentum</i>	Improved memory deficit	(61)

Dietary fiber

Digestive enzymes encoded in the human genome are unable to break down dietary fiber, which is defined as the carbohydrates present in plants (fruits, vegetables, legumes, nuts, seeds, and grains). Instead, only anaerobic fermentation by the gastrointestinal microbiota allows for the selective metabolism of dietary fiber (62). Dietary modifications impact the gut microbiome's composition, making it one of the most significant environmental influences. The different kinds of foods taken and the frequency of meals additionally have an impact on the synthesis of secondary bacterial metabolites because dietary components serve as substrates for gut germs (63). Which are primarily members of the *taxa Clostridium pneumonia* (phylum *Formicates*) and *Bacteriaemia* (phylum *Bacteroidetes*). Dietary fiber helps maintain community diversity in the mouse gut by altering the genetic makeup of the microbiome. Microbiota that rely exclusively on this vital resource are irreversibly destroyed by long-term low-fiber diets; even after decades of following high-fiber diets, they cannot be restored (64). Dietary fiber safeguards the intestinal barrier through SCFAs, specifically butyrate and acetate. Butyrate energizes enterocytes, maintaining gut epithelium integrity, while acetate reinforces barrier function by stimulating the interleukin-18 receptor, ensuring a healthy gut lining (65).

The Microbes Transplantation from feces

Fecal microbiota transplantation (FMT) involves transferring a processed stool sample from a donor into the gastrointestinal tract of a recipient, aiming to restore or improve the recipient's gut microbiota balance and promote health benefits. In the fourth century CE, Ge Hong described the use of stool as a treatment for a variety of diseases, especially gastroenteritis

(66). The use of excrement as medicine dates back to the earliest records of humanity. The debut of FMT into medical practice was signaled by Eiseman and the team's 1958 report on the use of stool bowel movements as a treatment for pseudomembranous colitis. The first step in the process is selecting an anonymous donor whose relatives have no family history of metabolic, autoimmune disorders, or cancerous process, and screening for potential infections (67). The next stage in preparing the feces is to mix them with water or regular saline, and then filter them to get rid of any particles. Various gastrointestinal access methods for combination therapy. A nasojejunal tube, nasogastric tube, esophagogastroduodenoscopy, a bowel examination, or retention enema can be used to administer the combination (68).

Donor Screening

To ensure the procedure's safety, the individual who donates must be properly chosen. Donor screening is vital for stopping the propagation of dangerous illnesses. In addition, there's a chance that FMT will alter an individual's susceptibility to diseases or diseases that impact the gastrointestinal flora. Before donation, every potential donor will have a confidential interview and tests in the laboratory, such as a blood and stool study, to reduce these risks (69).

Transplantation procedure

The two components of FMT are (1) digestive preparation and (2) decal material administration Using antibiotics to prepare the gut and create a habitat for the transplanted microorganisms to settle in and multiply is the first step in the FMT process (70). In contrast to either no preparation of the bowel or stool preparation with a laxative, pre-transplant antibiotic therapy permitted more effective engraftment in the mouse model of FMT, highlighting the significance of this(71). It may be more crucial when utilizing antibiotics to prepare the gut for FMT for purposes apart from CDI, as patients with CDI typically exhibit only a small number of microbes in their intestines. The procedure of transplanting fecal microbes involves two distinct stages. Before receiving a laxative, patients in the initial phase are given oral antibiotics to prepare their intestines. The recipient will be given the donor's stool sample. By upper or lower intestinal endoscopy, naso-enteral tubes, or capsule at least 24 hours after the last oral antibiotic treatment (72) (Figure8).

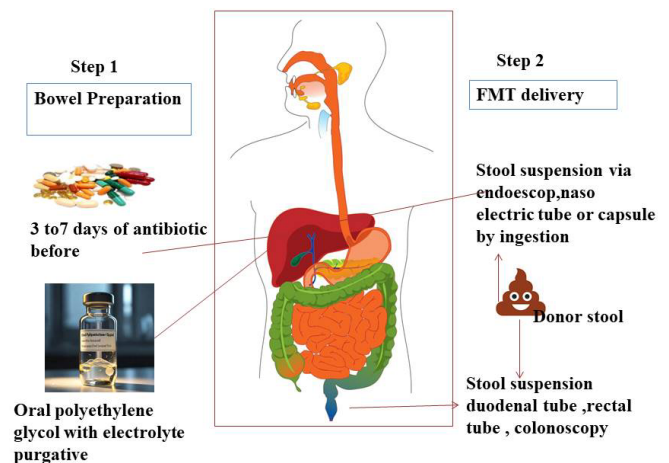


Figure 8. Fecal Transplantation Procedure

Mechanism Action of immune system regulation

1. T cell differentiation

AD, the most prevalent neurological condition worldwide, results in irreversible dementia and significant behavioural abnormalities. T-cells play an essential part in the immune pathogenesis of PD, AD, and MS in examination samples (73). While some T-cells (Th1, Th17, $\gamma\delta$ T-cells) turn on harmful microglia (M1-like), leading to neuron damage, other T-cells (Th2, Tregs, CD8+) promote neuroprotection by activating beneficial microglia (M2-like)(74). Neurodegeneration results from a concentration of M1-like microglia as the disease progresses. Yet during immune-suppressive treatment or remission, microglia shift to the protective M2-like phenotype (75). Research has linked Th1, Th17, and Treg cells to controlling neuroinflammation in PD and AD. Other T-cell subsets' roles remain unknown. In amyotrophic lateral sclerosis (ALS), T-cells regulate neuroinflammation. Encephalitogenic Tregs and Th2 cells naturally initiate anti-inflammatory responses, impeding neurodegeneration. Growth factors like IGF-1, BDNF, and GDNF support neuroprotection(75).

2. B Cell Activation

In autoimmune neurological disorders, B cells contribute to tissue damage through:

- Producing harmful antibodies that activate complement or trigger cell-mediated cytotoxicity.
- Presenting antigens, expanding cytotoxic T cells, and triggering cytokine release.
- Releasing proinflammatory cytokines (IL-6, TNF, GM-CSF) that activate macrophages.
- Forming ectopic lymphoid tissues, promoting local immune responses.

These functions exacerbate tissue damage and inflammation in autoimmune neurological disorders(76).

Metabolic production

a) Beneficial Metabolites:

Fatty acid SCFAs, obtained by the gut microbiota from resistant starch and dietary fiber, had a variety of influences on gut-brain communication SCFAs:

1. Include receptors linked to G-proteins (GPCRs) to regulate gut immunity and barrier function.
2. Induce gut hormone secretion (GLP1, PYY, GABA, 5-HT) via entering endocrine cells.
3. Reach systemic circulation, regulating liver function, insulin secretion, and energy homeostasis.
4. Cross the cerebral barrier, influencing integrity and neuroinflammation.
5. Influence glial cells, neurotropic factors, neurogenesis, and serotonin biosynthesis in the CNS.

SCFAs play a crucial role in maintaining gut-brain axis homeostasis, including overall health and neurological function(77).

b) Neurotransmitter Precursor

The impact of gut bacteria production on the inability to walk brain function. According to (78). the gut microbiota may

either catalyze the nutritional metabolism of neurotransmitters or synthesize neurotransmitter precursors. Certain kinds of bacteria can communicate by entering endocrine cells through their metabolites, showing the part gut bacteria play in paraplegics' brain function. Bacteria and enter endocrine cells can produce neurotransmitters that move through the bloodstream throughout the body. Certain precursors of neurotransmitters play a role in the brain's neurotransmitter synthesis process after crossing the BBB. Additionally, neuropod cells, which originate from the intestinal lining, generate and release neurotransmitters like glutamate. These cells can transmit sensory signals to the brain rapidly through the vagus nerve(79). Changes in the synthesis of transmitters as well as precursors that gut impact on brain health and neurological diseases. Abbreviations include L-DOPA GABA (gamma-aminobutyric acid), and 5-HTP (five-hydroxytryptophan)(80).

Gut Barrier integrity: (Figure 9)

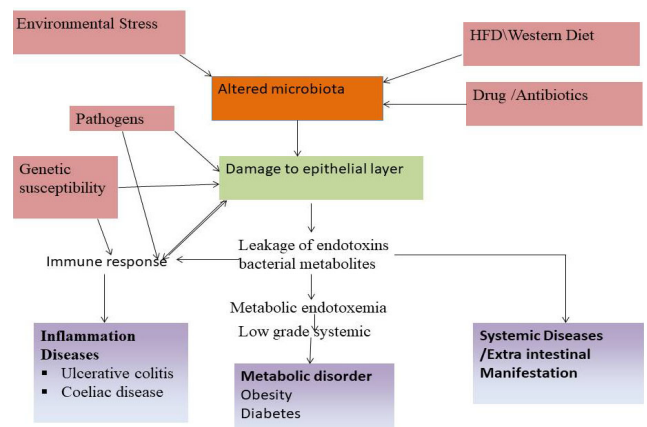


Figure 9. Factors affecting intestinal barrier integrity and pathological implications

Modifications in microbiota, inherited traits, nutrition, and an assortment of external variables can either directly or indirectly affect barrier integrity(81). Immune response is caused by compromised barrier integrity, which also contributes to an array of medical illness (82).

a) Tight junction modulation

Tight junctions are essential for the gut lining's health and leakiness. They encourage translocation across intestinal epithelia as an outcome (83). Pathogens employ multiple strategies with the goal of either directly disrupting the functional network or altering the various signaling pathways that play a role in modulating. Of these junctions next to a succinct overview of tight junction organization and modulation, we present the most recent findings regarding the molecular mechanisms causing the gut barrier to permeabilize as a result of pathogens such as bacteria, viruses, fungi, and parasites attacking tight junctions(84). Microbes disrupt tight junctions (TJs) through direct damage or receptor binding, triggering signaling pathways that compromise barrier function. This leads to inflammation, nitric oxide production, and permeability loss, facilitating infection(85).

Interleukin-10 (IL-10)

IL-10 is key to a healthy gut barrier integrity, protecting against damage from sepsis, lung injury, and total parenteral nutrition (TPN). Studies show IL-10:

- Prevents disruption of tight junctions (TJs) and adherens junctions
- Reduces intestinal permeability and bacterial translocation
- Reverses barrier defects
- Maintains indication of E-cadherin, ZO-1, claudin-1, and occluding

IL-10's barrier-protective effects highlight its importance in preventing intestinal damage and promoting epithelial health(86).

Mucus layer enhancement

The gut mucus layer serves a crucial part of retaining intestinal homeostasis and protecting the intestinal tract from chemical, mechanical, and biological threats. By coating the intestinal cells with a coat, it defends them from germs, digestive enzymes, and other harmful chemicals. The constant release of mucus roughly 10 L per day into the GI tract shows its significant protective role (87). The mucosal barrier's ability to resist infections depends on the degree to which it works in coordination with the immune system. Mucus is essential for preserving the innate intestinal barrier because it reduces the number of germs and antigens that get to the immune cells that live beneath the enterocytes. The body uses this as its main defense against chemicals that might be harmful(88,89).The mucus membrane appears to have a significant impact on immune function because the glycan moieties found in the mucus layer can interact directly with immune cells through lectin-like proteins. MUC2 mucin improves gut homeostasis and promotes oral tolerance by regulating the activity of intestinal epithelial cells and dendritic cells, whereas the MUC2 receptor complex has been demonstrated to reduce inflammatory responses in dendritic cells (90).

Limitations of studies involving gut microbiota and brain interactions

Although significant and specific investigations on the relationship between the microbiota of the gut and neurological diseases, a few holes persist, showing that an additional mechanistic grasp of how it works is required. The expression "gut dysbiosis" indicates and supports gut health, yet it seems ambiguous and overly general. Despite inter- and intra-individual variation in a niche, the animal and human microbes differ substantially in along with inter- and intra-individual fluctuations in a niche, the animal and human bacteria differ considerably in.

REFERENCES

1. Carloni S, Rescigno M. The gut-brain vascular axis in neuroinflammation. In *Seminars in immunology* 2023 Sep 1 (Vol. 69, p. 101802). Academic Press.
2. Kandpal M, Indari O, Baral B, Jakhmola S, Tiwari D, Bhandari V, Pandey RK, Bala K, Sonawane A, Jha HC. Dysbiosis of gut microbiota from the perspective of the gut-brain axis: role in the provocation of neurological disorders. *Metabolites*. 2022 Nov 3;12(11):1064.
3. Leclercq S, Matamoros S, Cani PD, Neyrinck AM, Jamar F, Stärkel P, Windey K, Tremaroli V, Bäckhed F, Verbeke K, de Timary P. Intestinal permeability, gut-bacterial dysbiosis, and behavioral markers of alcohol-dependence severity. *Proceedings of the National Academy of Sciences*. 2014 Oct 21;111(42):E4485-93.
4. Mayer, E. A., Knight, R., Mazmanian, S. K., Cryan, J. F., & Tillisch, K. (2014). Gut microbes and the brain: a paradigm shift in neuroscience. *Journal of Neuroscience*, 34(46), 15490-15496.
5. Liang J, Liu B, Dong X, Wang Y, Cai W, Zhang N, Zhang H. Decoding the role of gut microbiota in Alzheimer's pathogenesis and envisioning future therapeutic avenues. *Frontiers in Neuroscience*. 2023 Sep 18;17:1242254.
6. Pourahmad, R., Gholinejad, M. Z., Aram, C., Farsani, A. S., Banazadeh, M., & Tafakhori, A. (2024). Exploring the effect of gut microbiome on Alzheimer's disease. *Biochemistry and Biophysics Reports*, 39, 101776.
7. Martini GA, Phear EA, Ruebner B, Sherlock S. The bacterial content of the small intestine in normal and cirrhotic subjects: relation to methionine toxicity.
8. Phear EA, Ruebner B, Sherlock S, Summerskill WH. Methionine toxicity in liver disease and its prevention by chlortetracycline.
9. Foster JA, Lyte M, Meyer E, Cryan JF. Gut microbiota and brain function: an evolving field in neuroscience. *International Journal of Neuropsychopharmacology*. 2016 May 1;19(5):pyv114.
10. Oroojzadeh P, Bostanabad SY, Lotfi H. Psychobiotics: the influence of gut microbiota on the gut-brain axis in neurological disorders. *Journal of Molecular Neuroscience*. 2022 Sep;72(9):1952-64.
11. Goyal D, Ali SA, Singh RK. Emerging role of gut microbiota in modulation of neuroinflammation and neurodegeneration with emphasis on Alzheimer's disease. *Progress in Neuro-Psychopharmacology and Biological Psychiatry*. 2021 Mar 2;106:110112.
12. Raval U, Harary JM, Zeng E, Pasinetti GM. The dichotomous role of the gut microbiome in exacerbating and ameliorating neurodegenerative disorders. *Expert review of neurotherapeutics*. 2020 Jul 2;20(7):673-86.
13. Xin SH, Tan L, Cao X, Yu JT, Tan L. Clearance of amyloid beta and tau in Alzheimer's disease: from mechanisms to therapy. *Neurotoxicity research*. 2018 Oct;34:733-48.
14. Wang C, Klechikov AG, Gharibyan AL, Wärmländer SK, Jarvet J, Zhao L, Jia X, Shankar SK, Olofsson A, Brännström T, Mu Y. The role of pro-inflammatory S100A9 in Alzheimer's disease amyloid-neuroinflammatory cascade. *Acta neuropathologica*. 2014 Apr;127:507-22.
15. Canfora EE, Jocken JW, Blaak EE. Short-chain fatty acids in control of body weight and insulin sensitivity. *Nature Reviews Endocrinology*. 2015 Oct;11(10):577-91.
16. Dinan TG, Cryan JF. Gut instincts: microbiota as a key regulator of brain development, ageing and neurodegeneration. *The Journal of physiology*. 2017 Jan 15;595(2):489-503.
17. Wang S, Mims PN, Roman RJ, Fan F. Is beta-amyloid accumulation a cause or consequence of Alzheimer's disease?. *Journal of Alzheimer's parkinsonism & dementia*. 2016 Nov 17;1(2):007.
18. Bourassa MW, Alim I, Bultman SJ, Ratan RR. Butyrate, neuroepigenetics and the gut microbiome: can a high fiber diet improve brain health?. *Neuroscience letters*. 2016 Jun 20;625:56-63.
19. Kowalski K, Mulak A. Brain-gut-microbiota axis in Alzheimer's disease. *Journal of neurogastroenterology and motility*. 2019 Jan 1;25(1):48.

20. Mangialasche F, Solomon A, Winblad B, Mecocci P, Kivipelto M. Alzheimer's disease: clinical trials and drug development. *The Lancet Neurology*. 2010 Jul 1;9(7):702-16..
21. Mullane K, Williams M. Alzheimer's therapeutics: continued clinical failures question the validity of the amyloid hypothesis—but what lies beyond?. *Biochemical pharmacology*. 2013 Feb 1;85(3):289-305..
22. Holmes C, Boche D, Wilkinson D, Yadegarfar G, Hopkins V, Bayer A, Jones RW, Bullock R, Love S, Neal JW, Zotova E. Long-term effects of A β 42 immunisation in Alzheimer's disease: follow-up of a randomised, placebo-controlled phase I trial. *The Lancet*. 2008 Jul 19;372(9634):216-23.
23. Korte N, Nortley R, Attwell D. Cerebral blood flow decrease as an early pathological mechanism in Alzheimer's disease. *Acta neuropathologica*. 2020 Dec;140(6):793-810..
24. Barnham KJ, Masters CL, Bush AI. Neurodegenerative diseases and oxidative stress. *Nature reviews Drug discovery*. 2004 Mar 1;3(3):205-14.
25. Bosco DA, Fowler DM, Zhang Q, Nieva J, Powers ET, Wentworth P, Lerner RA, Kelly JW. Elevated levels of oxidized cholesterol metabolites in Lewy body disease brains accelerate α -synuclein fibrilization. *Nature chemical biology*. 2006 May;2(5):249-53.
26. Breydo L, Wu JW, Uversky VN. α -Synuclein misfolding and Parkinson's disease. *Biochimica et Biophysica Acta (BBA)-Molecular Basis of Disease*. 2012 Feb 1;1822(2):261-85.
27. Sampson TR, Debelius JW, Thron T, Janssen S, Shastri GG, Ilhan ZE, Challis C, Schretter CE, Rocha S, Gradinaru V, Chesselet MF. Gut microbiota regulate motor deficits and neuroinflammation in a model of Parkinson's disease. *Cell*. 2016 Dec 1;167(6):1469-80.
28. Houser MC, Tansey MG. The gut-brain axis: is intestinal inflammation a silent driver of Parkinson's disease pathogenesis?. *NPJ Parkinson's disease*. 2017 Jan 11;3(1):3.
29. De Theije CG, Wopereis H, Ramadan M, van Eijndthoven T, Lambert J, Knol J, Garssen J, Kraneveld AD, Oozeer R. Altered gut microbiota and activity in a murine model of autism spectrum disorders. *Brain, behavior, and immunity*. 2014 Mar 1;37:197-206.
30. Sun MF, Shen YQ. Dysbiosis of gut microbiota and microbial metabolites in Parkinson's Disease. *Ageing research reviews*. 2018 Aug 1;45:53-61.
31. Keshavarzian A, Green SJ, Engen PA, Voigt RM, Naqib A, Forsyth CB, Mutlu E, Shannon KM. Colonic bacterial composition in Parkinson's disease. *Movement Disorders*. 2015 Sep;30(10):1351-60.
32. Gray MT, Woulfe JM. Striatal blood-brain barrier permeability in Parkinson's disease. *Journal of Cerebral Blood Flow & Metabolism*. 2015 May;35(5):747-50.
33. Fazekas F, Enzinger C, Wallner-Blazek M, Ropele S, Pluta-Fuerst A, Fuchs S. Gender differences in MRI studies on multiple sclerosis. *Journal of the neurological sciences*. 2009 Nov 15;286(1-2):28-30.
34. Trojano M, Lucchese G, Graziano G, Taylor BV, Simpson Jr S, Lepore V, Grand'Maison F, Duquette P, Izquierdo G, Grammond P, Amato MP. Geographical variations in sex ratio trends over time in multiple sclerosis. *PloS one*. 2012 Oct 25;7(10):e48078.
35. Coles A. Multiple sclerosis: the bare essentials. *Practical neurology*. 2009 Apr 1;9(2):118-26.
36. Nicol B, Salou M, Laplaud DA, Wekerle H. The autoimmune concept of multiple sclerosis. *La Presse Médicale*. 2015 Apr 1;44(4):e103-12.
37. Nicol B, Salou M, Laplaud DA, Wekerle H. The autoimmune concept of multiple sclerosis. *La Presse Médicale*. 2015 Apr 1;44(4):e103-12.
38. Stockinger B, Veldhoen M, Martin B. Th17 T cells: linking innate and adaptive immunity. In *Seminars in immunology* 2007 Dec 1 (Vol. 19, No. 6, pp. 353-361). Academic Press.
39. Obermeier B, Lovato L, Mentele R, Brück W, Forne I, Imhof A, Lottspeich F, Turk KW, Willis SN, Wekerle H, Hohlfeld R. Related B cell clones that populate the CSF and CNS of patients with multiple sclerosis produce CSF immunoglobulin. *Journal of neuroimmunology*. 2011 Apr 1;233(1-2):245-8..
40. Yan W, Nguyen T, Yuki N, Ji Q, Yiannikas C, Pollard JD, Mathey EK. Antibodies to neurofascin exacerbate adoptive transfer experimental autoimmune neuritis. *Journal of Neuroimmunology*. 2014 Dec 15;277(1-2):13-7.
41. Lebeer S, Vanderleyden J, De Keersmaecker SC. Host interactions of probiotic bacterial surface molecules: comparison with commensals and pathogens. *Nature Reviews Microbiology*. 2010 Mar;8(3):171-84.
42. Lieberman SM, Evans AM, Han B, Takaki T, Vinnitskaya Y, Caldwell JA, Serreze DV, Shabanowitz J, Hunt DF, Nathenson SG, Santamaria P. Identification of the β cell antigen targeted by a prevalent population of pathogenic CD8 $^{+}$ T cells in autoimmune diabetes. *Proceedings of the National Academy of Sciences*. 2003 Jul 8;100(14):8384-8.
43. Buscarinu MC, Cerasoli B, Annibali V, Policano C, Lionetto L, Capi M, Mechelli R, Romano S, Fornasiero A, Mattei G, Piras E. Altered intestinal permeability in patients with relapsing-remitting multiple sclerosis: A pilot study. *Multiple Sclerosis Journal*. 2017 Mar;23(3):442-6.
44. Tarawneh R, Penhos E. The gut microbiome and Alzheimer's disease: Complex and bidirectional interactions. *Neuroscience & Biobehavioral Reviews*. 2022 Oct 1;141:104814.
45. Diaz-Marugan L, Kantsjö JB, Rutsch A, Ronchi F. Microbiota, diet, and the gut-brain axis in multiple sclerosis and stroke. *European Journal of Immunology*. 2023 Nov;53(11):2250229.
46. Gloor SM, Wachtel M, Bolliger MF, Ishihara H, Landmann R, Frei K. Molecular and cellular permeability control at the blood-brain barrier. *Brain research reviews*. 2001 Oct 1;36(2-3):258-64.
47. Profaci CP, Munji RN, Pulido RS, Daneman R. The blood-brain barrier in health and disease: Important unanswered questions. *Journal of Experimental Medicine*. 2020 Mar 25;217(4):e20190062.
48. Rastogi S, Singh A. Gut microbiome and human health: Exploring how the probiotic genus *Lactobacillus* modulate immune responses. *Frontiers in Pharmacology*. 2022 Oct 24;13:1042189.
49. Zhou H, Wang R, Zhang S, Zhang X, Zhou H, Wen T, Wang J. Depression-like symptoms due to *Defl* deficiency are alleviated by intestinal transplantation of *Lactobacillus murine* and *Lactobacillus reuteri*. *Biochemical and Biophysical Research Communications*. 2022 Feb 19;593:137-43.
50. Joung H, Chu J, Kim BK, Choi IS, Kim W, Park TS. Probiotics ameliorate chronic low-grade inflammation and fat accumulation with gut microbiota composition change in diet-induced obese mice models. *Applied Microbiology and Biotechnology*. 2021 Feb;105:1203-13.

51. Verma R, Lee C, Jeun EJ, Yi J, Kim KS, Ghosh A, Byun S, Lee CG, Kang HJ, Kim GC, Jun CD. Cell surface polysaccharides of *Bifidobacterium bifidum* induce the generation of Foxp3+ regulatory T cells. *Science immunology*. 2018 Oct 19;3(28):eaat6975.
52. Henrick BM, Rodriguez L, Lakshmikanth T, Pou C, Henckel E, Arzoomand A, Olin A, Wang J, Mikes J, Tan Z, Chen Y. Bifidobacteria-mediated immune system imprinting early in life. *Cell*. 2021 Jul 22;184(15):3884-98..
53. Henrick BM, Rodriguez L, Lakshmikanth T, Pou C, Henckel E, Arzoomand A, Olin A, Wang J, Mikes J, Tan Z, Chen Y. Bifidobacteria-mediated immune system imprinting early in life. *Cell*. 2021 Jul 22;184(15):3884-98.
54. Lim HJ, Shin HS. Antimicrobial and immunomodulatory effects of bifidobacterium strains: A review. *Journal of Microbiology and Biotechnology*. 2020 Oct 20;30(12):1793.
55. Ikeda Y, Taniguchi K, Yoshikawa S, Sawamura H, Tsuji A, Matsuda S. A budding concept with certain microbiota, anti-proliferative family proteins, and engram theory for the innovative treatment of colon cancer. *Exploration of Medicine*. 2022 Oct 27;3(5):468-78.
56. Den H, Dong X, Chen M, Zou Z. Efficacy of probiotics on cognition, and biomarkers of inflammation and oxidative stress in adults with Alzheimer's disease or mild cognitive impairment—A meta-analysis of randomized controlled trials. *Aging (Albany NY)*. 2020 Feb 15;12(4):4010.
57. Kobayashi Y, Sugahara H, Shimada K, Mitsuyama E, Kuhara T, Yasuoka A, Kondo T, Abe K, Xiao JZ. Therapeutic potential of *Bifidobacterium breve* strain A1 for preventing cognitive impairment in Alzheimer's disease. *Scientific reports*. 2017 Oct 18;7(1):13510.
58. Athari Nik Azm S, Djazayeri A, Safa M, Azami K, Ahmadvand B, Sabbaghziarani F, Sharifzadeh M, Vafa M. Lactobacilli and bifidobacteria ameliorate memory and learning deficits and oxidative stress in β -amyloid (1–42) injected rats. *Applied Physiology, Nutrition, and Metabolism*. 2018;43(7):718-26.
59. Ayten Ş, Bilici S. Modulation of gut microbiota through dietary intervention in neuroinflammation and Alzheimer's and Parkinson's diseases. *Current Nutrition Reports*. 2024 Jun;13(2):82-96.
60. Slavin J. Impact of the proposed definition of dietary fiber on nutrient databases. *Journal of Food Composition and Analysis*. 2003 Jun 1;16(3):287-91.
61. Reynolds A, Mann J, Cummings J, Winter N, Mete E, TeMorenga L. Carbohydrate quality and human health: a series of systematic reviews and meta-analyses. *The Lancet*. 2019 Feb 2;393(10170):434-45.
62. Usuda H, Okamoto T, Wada K. Leaky gut: effect of dietary fiber and fats on microbiome and intestinal barrier. *International journal of molecular sciences*. 2021 Jul 16;22(14):7613.
63. Bakken JS, Borody T, Brandt LJ, Brill JV, Demarco DC, Franzos MA, Kelly C, Khoruts A, Louie T, Martinelli LP, Moore TA. Treating *Clostridium difficile* infection with fecal microbiota transplantation. *Clinical gastroenterology and hepatology*. 2011 Dec 1;9(12):1044-9.
64. Zhang F, Luo W, Shi Y, Fan Z, Ji G. Should we standardize the 1,700-year-old fecal microbiota transplantation?. *American Journal of Gastroenterology (Springer Nature)*. 2012 Nov 1;107(11).
65. EISEMAN B, Silen W, Gs B, Aj K. Fecal enema as an adjunct in the treatment of pseudomembranous enterocolitis. *Surgery*. 1958 Nov 1;44(5):854-9.
66. Smits LP, Bouter KE, De Vos WM, Borody TJ, Nieuwdorp M. Therapeutic potential of fecal microbiota transplantation. *Gastroenterology*. 2013 Nov 1;145(5):946-53.
67. Arajol C, Gómez AA, Gonzalez-Suarez B, Casals-Pascual C, Martí SM, Luzón MÁ, Soriano A, Capón JG. Donor selection for faecal microbiota transplantation. Consensus document of the Catalan Society of Gastroenterology and the Catalan Society of Infectious Diseases and Clinical Microbiology. *Gastroenterología Y Hepatología (English Edition)*. 2021 Feb 1;44(2):175-80.
68. Bou Zerdan M, Niforatos S, Nasr S, Nasr D, Ombada M, John S, Dutta D, Lim SH. Fecal microbiota transplant for hematologic and oncologic diseases: principle and practice. *Cancers*. 2022 Jan 29;14(3):691.
69. Fecal Microbiota Transplantation-standardization Study Group. Nanjing consensus on methodology of washed microbiota transplantation. *Chinese medical journal*. 2020 Oct 5;133(19):2330-2.
70. Peng Z, Xiang J, He Z, Zhang T, Xu L, Cui B, Li P, Huang G, Ji G, Nie Y, Wu K. Colonic transendoscopic enteral tubing: a novel way of transplanting fecal microbiota. *Endoscopy international open*. 2016 Jun;4(06):E610-3.
71. Lucin KM, Wyss-Coray T. Immune activation in brain aging and neurodegeneration: too much or too little?. *Neuron*. 2009 Oct 15;64(1):110-22.
72. Brochard V, Combadière B, Prigent A, Laouar Y, Perrin A, Beray-Berthet V, Bonduelle O, Alvarez-Fischer D, Callebert J, Launay JM, Duyckaerts C. Infiltration of CD4+ lymphocytes into the brain contributes to neurodegeneration in a mouse model of Parkinson disease. *The Journal of clinical investigation*. 2008 Dec 22;119(1).
73. Harms AS, Cao S, Rowse AL, Thome AD, Li X, Mangieri LR, Cron RQ, Shacka JJ, Raman C, Standaert DG. MHCII is required for α -synuclein-induced activation of microglia, CD4 T cell proliferation, and dopaminergic neurodegeneration. *Journal of Neuroscience*. 2013 Jun 5;33(23):9592-600.
74. González H, Contreras F, Prado C, Elgueta D, Franz D, Bernales S, Pacheco R. Dopamine receptor D3 expressed on CD4+ T cells favors neurodegeneration of dopaminergic neurons during Parkinson's disease. *The Journal of Immunology*. 2013 May 15;190(10):5048-56.
75. Stathopoulos P, Dalakas MC. Evolution of anti-B cell therapeutics in autoimmune neurological diseases. *Neurotherapeutics*. 2022 Apr 1;19(3):691-710.
76. Silva YP, Bernardi A, Frozza RL. The role of short-chain fatty acids from gut microbiota in gut-brain communication. *Frontiers in endocrinology*. 2020 Jan 31;11:508738.
77. Caspani G, Swann J. Small talk: microbial metabolites involved in the signaling from microbiota to brain. *Current opinion in pharmacology*. 2019 Oct 1;48:99-106.
78. Jiang Y, Li K, Li X, Xu L, Yang Z. Sodium butyrate ameliorates the impairment of synaptic plasticity by inhibiting the neuroinflammation in 5XFAD mice. *Chemico-biological interactions*. 2021 May 25;341:109452.
79. Kaelberer MM, Buchanan KL, Klein ME, Barth BB, Montoya MM, Shen X, Bohórquez DV. A gut-brain neural circuit for nutrient sensory transduction. *Science*. 2018 Sep 21;361(6408):eaat5236.

80. Ma TY. Intestinal epithelial barrier dysfunction in Crohn's disease. *Proceedings of the Society for Experimental Biology and Medicine*. 1997 Apr;214(4):318-27.
81. Chelakkot C, Ghim J, Ryu SH. Mechanisms regulating intestinal barrier integrity and its pathological implications. *Experimental & molecular medicine*. 2018 Aug;50(8):1-9.
82. Paradis T, Bègue H, Basmaciyan L, Dalle F, Bon F. Tight junctions as a key for pathogens invasion in intestinal epithelial cells. *International journal of molecular sciences*. 2021 Mar 2;22(5):2506.
83. Mu Q, Kirby J, Reilly CM, Luo XM. Leaky gut as a danger signal for autoimmune diseases. *Frontiers in immunology*. 2017 May 23;8:269575.
84. Guttman JA, Finlay BB. Tight junctions as targets of infectious agents. *Biochimica et Biophysica Acta (BBA)-Biomembranes*. 2009 Apr 1;1788(4):832-41.
85. Wang Q, Hasselgren PO. Heat shock response reduces intestinal permeability in septic mice: potential role of interleukin-10. *American Journal of Physiology-Regulatory, Integrative and Comparative Physiology*. 2002 Mar 1;282(3):R669-76.
86. Cone RA. Barrier properties of mucus. *Advanced drug delivery reviews*. 2009 Feb 27;61(2):75-85.
87. Cornick S, Tawiah A, Chadee K. Roles and regulation of the mucus barrier in the gut. *Tissue barriers*. 2015 Apr 3;3(1-2):e982426.
88. Johansson ME, Sjövall H, Hansson GC. The gastrointestinal mucus system in health and disease. *Nature reviews Gastroenterology & hepatology*. 2013 Jun;10(6):352-61.
89. Pelaseyed T, Bergström JH, Gustafsson JK, Ermund A, Birchenough GM, Schütte A, van der Post S, Svensson F, Rodríguez-Piñeiro AM, Nyström EE, Wising C. The mucus and mucins of the goblet cells and enterocytes provide the first defense line of the gastrointestinal tract and interact with the immune system. *Immunological reviews*. 2014 Jul;260(1):8-20.
90. Corfield AP. Mucins: a biologically relevant glycan barrier in mucosal protection. *Biochimica et Biophysica Acta (BBA)-General Subjects*. 2015 Jan 1;1850(1):236-52.
